Study of in-vitro biotransformation of prodrugs of Ester and Amide linkages of Ibuprofen, Diclofenac Sodium and Paracetamol in acidic and alkaline medium

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ABSTRACT

Our goal is to perform in-vitro biotransformation of Prodrugs of ibuprofen+paracetamol, diclofenac sodium+paracetamol and ibuprofen+diclofenac sodium by acidic and alkaline hydrolysis of both ester (-COO-) and amide (-CONH-) linkages into free drugs and chromatographically separation of their Rt in HPLC. Since both ester (-COO-) and amide (-CONH-) linkages are susceptible to hydrolysis in both acidic pH (gastric pH) and basic pH (intestinal pH) to produce parent drug ibuprofen, diclofenac and paracetamol by biotransformation in in-vivo; so it will be implemented as a Prodrug which can show prolong action on pain and fever after getting released into free parent drug by biotransformation. The HPLC (High-Performance Liquid Chromatography) study reports the retention time (Rt) and releases kinetics of three Prodrugs by taking HPLC degradation data of three samples of Prodrugs and individual HPLC data of parent drugs separately to compare the Rt value of release of three drugs from Prodrugs in both acidic and alkaline pH. Prodrug–A (logP=4.56) releases lbuprofen & Paracetamol, Prodrug–B (logP=4.90) releases Diclofenac & Paracetamol and Prodrug–C (logP=6.13) releases lbuprofen & Diclofenac. This is a comparison study of drug release in in-vitro gastric as well as intestinal pH focusing on in-vivo biotransformation.

Keywords: Prodrug-A/Prodrug-B/Prodrug-C, Ibuprofen, Paracetamol, Diclofenac sodium, Molecular weight, logP, UV λmax, IR, Mobile phase, TLC-Rf value, HPLC-Rt value, LOD, LOC

INTRODUCTION

Three Prodrugs (Prodrug–A, Prodrug–B and Prodrug–C) have been successfully synthesized and have shown different melting points from individual parent drugs (Ibuprofen, Diclofenac sodium, and Paracetamol) which indicate the authenticity of the fulfillment of Prodrug synthesis. ^[1,2]



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The synthesized Prodrugs were characterized by m.p., log P values and IR (Infra Red) spectrum for structural identification. Their solubility parameters also found different from parent drugs. A prodrug is a substance having no medicinal importance but after biotransformation in GIT, it releases the parent drug which is able to show the pharmacological activity.^[3-5]

Ibuprofen [$C_{13}H_{18}O_2$; MW=206] (2–[4–(2–methylpropyl)phenyl]propanoic acid), from isobutylphenylpropanoic acid, is a drug in the nonsteroidal anti–inflammatory drug (NSAID) class used for treating pain, fever and inflammation. logP=3.97, pKa=4.91. Ibuprofen m.p.=theoretical (**76°C**), Practical (**75°C**).

Diclofenac sodium [NaC₁₄H₁₀Cl₂NO; MW=302] (Sodium {2–[(2,6–dichlorophenyl)amino]phenyl}acetic acetate) is a nonsteroidal anti–inflammatory drug (NSAID) taken or applied to reduce inflammation and as an analgesic reducing pain in certain conditions. logP=4.51, pKa=4.15. Diclofenac sodium m.p.=theoretical (**283–285°C**), Practical (**282°C**)

Paracetamol [$C_8H_9NO_2$; MW=151] (N–(4–hydroxyphenyl)acetamide), also known as acetaminophen or APAP, is a medication used to treat pain and fever. logP=0.46, pKa=9.38. Paracetamol m.p.=theoretical (**169°C**), Practical (**170°C**).

Experimental part:

The mobile phase of TLC values has been selected after trial and error for all individual NSAID (ibuprofen, diclofenac sodium and paracetamol) according to their logP and R_f values have been determined and after that R_f values have been obtained for all three Prodrugs (Prodrug-A, Prodrug-B, Prodrug-C). The mobile phase solvent ratio of TLC helped to select the mobile phase selection for HPLC.^[6-8] The values are as follows:



UV maximum Absorbance (Prodrug-A)



UV maximum Absorbance (Prodrug-C) Figure-2: λ_{max} of Prodrugs

Samples	Molecular Formula	% Yield	Melting Point (°C)	Molecular Weight	λ _{max} (nm)	logP						
Prodrug-A	$C_{21}H_{25}NO_{3}$	83.67	152–155°C	339	275	4.56						
Prodrug-B	$C_{22}H_{18}CI_2N_2O_3$	88.65	220–222°C	429	262	4.90						
Prodrug-C	$C_{27}H_{27}CI_2NO_3$	91.32	100–105°C	484	276	6.13						

Table-1: Physicochemical parameters of Prodrugs

Solubility study of Prodrug-A, Prodrug-B and Prodrug-C:

The solubility of Prodrug-A, Prodrug-B and Prodrug-C were practically determined. Solubility was determined by taking 100mg of Prodrug-A, Prodrug-B and Prodrug-C respectively in a 10ml volumetric flask, adding required quantity of solvent at room temperature and shaken for few minutes.^[9-11] Solubility data for each study was observed and recorded in Table-2.

Solvent	Solubility									
Water	Prodrug-A: Soluble, Prodrug-B: Insoluble, Prodrug-C: Slightly soluble									
Methanol	Freely soluble									
Acetone	Freely soluble									

Table-2: Solubility parameters of Prodrugs

Instrument:

A double beam UV visible spectrophotometer: Manufacturer: Shimadzu. Model: UV-1800, Shimadzu, Japan.

Preparation of Standard solution of Prodrug-A: Weighed accurately about 100mg of Prodrug-A separately transferred quantitatively to the 100ml volumetric flask. Dissolved in about 70ml of Methanol by sonication and diluted to volume with methanol and mixed. Transferred 0.1ml of this solution to the 10ml volumetric flask,

diluted to volume with diluents (methanol) and mixed ($10\mu g/ml$) and absorption was observed at 275 nm.^[12] The values of three Prodrugs were given in the Table-3.

	Conc. (µg/ml)	Abs.		Conc. (µg/ml)	Abs.		Conc. (µg/ml)	Abs.
Prodrug-A	10	0.112	Prodrug-B	10	0.208	Prodrug-C	10	0.441
	20	0.222		20	0.438		20	0.945
	30	0.338		30	0.664		30	1.216
	40	0.483		40	0.845		40	1.536

Table-3: Absorption data of Prodrug-A, Prodrug-B and Prodrug-C

Infra Red Spectral studies of Prodrug-A, Prodrug-B and Prodrug-C:

IR spectra of three Prodrugs were measured in KBr pellets in Shimadzu FT-IR spectrophotometer and values in cm⁻¹ were obtained to the interpretation of structural framework.



Figure-3: IR Spectra of Prodrug-A

Prodrug-A: IR (cm⁻¹; v): CH₃ asym. stretch: (standard=2970-2950/2880-2860, found=2953.45), >C=O stretching: (standard=1725-1705, found=1714.41), Amide II band: (standard=1680-1630, found=1509.03), CH-CO deformation: (standard=1470-1430/1380-1370, found=1420.32), C=C stretching: (standard=1615–1580, found=1610), C-N-H group: (standard=1090–1020, found=1070.3), Para-disubstituted aromatic ring: (standard=860–800, found=850).^[13]



Figure-4: IR Spectra of Prodrug-B

Prodrug-B: IR (cm⁻¹; ν): N-H stretching: (standard=3500-3100, found=3343), C-H stretching (aromatic): (standard=3150-3050, found=3136.36), C=C stretching: (standard=1615–1580, found=1621), C-N stretching: (standard=1650–1550, found=1580), >C=O stretching: (standard=1725-1705, found=1721), C-CO-C stretching: (standard=1320-1210, found=1284.63).^[14]



Figure-5: IR Spectra of Prodrug-C

Prodrug-C: IR (cm⁻¹; v): Amide (–CONH–): (standard=1630–1680, found=1735), Ketone (>C=O): (standard=1705–1725, found=1735), Imino (–NH–): (standard: 3350–3320, found=3350), Chlorine (–Cl): (standard=700-800, found=747, 778), Phenyl (–C₆H₅): (standard=1615–1580, found=1577), Carboxylic acid (–COOH): (standard=1700–1725, found=1734), Methyl (CH₃–): (standard=2970–2950/2880–2860, found=2952) Methyl C– H asym./sym. stretch standard=1470–1430/1380–1370, found=1447).^[15]

DEVELOPMENT AND VALIDATION OF STABILITY INDICATING RP-HPLC METHOD FOR PRODRUG-A (IBUPROFEN AND PARACETAMOL)

Experimental: Reagents and Materials, Prodrug-A synthesized in our college lab, Methanol (HPLC grade, Finar Chemicals Ltd, Ahmedabad, India), Water (HPLC grade, Finar Chemicals Ltd, Ahmedabad, India).

Equipment and Instruments: Shimadzu HPLC instrument (LC-2010 CHT) equipped with prominence diode array detector (SPD-M20A) (Software LC Solution), Analytical balance (Acculab ALC-2014, Huntingdon Valley, PA), Ultra sonicator (EN 30 US, Enertech Fast Clean, Mumbai, India), Hot air oven (TO-90S, Thermolab, Mumbai, India), pH meter (Thermo Electron Corp., Pune, India)

Development and Optimization of RP-HPLC Method:

a) Selection of Wavelength: The sensitivity of HPLC method that used UV detection depends upon proper selection of detection wavelength. An ideal wavelength is the one that gives good response for the drugs that are to be detected. The λ_{max} of Prodrug-A was 275nm in methanol, Prodrug-B was 262nm in methanol and Prodrug-C was 276nm in methanol.^[16]

b) Selection of Chromatographic Conditions: Proper selection of the HPLC method depends upon the nature of the sample (ionic or ionisable or neutral molecule), its molecular weight, pKa and solubility. RP-HPLC was selected for the initial separation based on literature survey and its simplicity and suitability. To optimize the

chromatographic conditions the effects of chromatographic variables such as mobile phase, pH, flow rate and solvent ratio were studied and the chromatographic parameters such as capacity factor, asymmetric factor, resolution and column efficiency were calculated. The pH of gastric acid varies from 1.5-3.5 in the human stomach lumen, the acidity being maintained by the proton pump H^*/K^* ATPase. So the pattern for acid hydrolysis was adjusted at pH=3-3.5 by HCl. The pH of intestine varies from 5.6-6.9, so the pattern for alkaline hydrolysis was adjusted at pH=7.0-8.0 by NaOH. Finally, the condition was chosen that gave the best resolution, symmetry and capacity factor was selected for estimation of Prodrug-A, Prodrug-B and Prodrug-C.^[17]

c) Selection of Ratio of Mobile phase: The solution containing 100µg/ml of Prodrug-A, Prodrug-B and Prodrug-C respectively was chromatographed with a mobile phase of different ratio of methanol and water.^[18]

	Trials	Ratio	Remark		Trials	Ratio	Remark		Trials	Ratio	Remark
	1	Methanol: Water (60:40)	Tailing		1	ACN: Water (80:20)	Tailing		1	Methano I: Water (90:10)	Tailing
	2	Methanol: Water (70:30)	Tailing		2	ACN: Water (70:30)	Tailing		2	Methano I: Water (85:15)	Tailing
Prodrug-A	3	ACN: Water (60:40)	Tailing	Prodrug-B	3	ACN: Methano I (80:20)	Tailing	Prodrug-C	3	Methano I: Water (70:30)	Tailing
	4	ACN: Water (70:30)	Tailing		4	ACN: Methano I (70:30)	Tailing		4	Methano I: Water (80:20)	Symmetrical peak
	5	Methanol: Water (80:20)	Symmetrical peak		5	Methano I: Water (80:20)	Tailing				
				6	Methano l: Water (70:30)	Symmetrical peak					

Table-4: Selection of mobile phase for Prodrug-A, Prodrug-B and Prodrug-C

PROCEDURE FOR SOLUTION PREPARATION FOR PRODRUG-A

1A. Preparation of synthesized drug stock solution: Accurately weighed 100mg (0.1gm) of Prodrug-A was transferred into 100ml volumetric flask, dissolved and diluted up to mark with diluents to yield a stock solution having a concentration of 1000µg/ml Prodrug-A.^[19]

2A. Preparation of working solution containing Prodrug-A: From the above solution (1000µg/ml of Prodrug-A) 0.2, 0.4, 0.6, 0.8, 1.0, 1.2ml was taken into 10ml volumetric flask to get concentration 20, 40, 60, 80, 100, 120µg/ml of Prodrug-A respectively.^[20]

PROCEDURE FOR SOLUTION PREPARATION FOR PRODRUG-B

1B. Preparation of synthesized drug stock solution: Accurately weighed 100mg (0.1gm) of Prodrug-B was transferred into 100ml volumetric flask, dissolved and diluted up to mark with diluents to yield a stock solution having a concentration of 1000μ g/ml Prodrug-B.^[21]

2B. Preparation of working solution containing Prodrug-B: From the above solution (1000μ g/ml of Prodrug-B), 0.01, 0.05, 0.10, 0.15, 0.20, 0.25ml was taken in to10ml volumetric flask to get concentration 1, 5, 10, 15, 20, 25 μ g/ml of Prodrug-B respectively.^[22]

PROCEDURE FOR SOLUTION PREPARATION FOR PRODRUG-C

1C. Preparation of synthesized drug stock solution: Accurately weighed 100mg (0.1gm) of Prodrug-C was transferred into 100ml volumetric flask, dissolved and diluted up to mark with diluents to yield a stock solution having a concentration of 1000µg/ml Prodrug-C.^[23]

2C. Preparation of working solution containing Prodrug-C: From the above solution (1000μ g/ml of Prodrug-C), 0.1, 0.15, 0.2, 0.25, 0.3, 0.35ml was taken in to 10ml volumetric flask to get concentration 10, 15, 20, 25, 30, 35μ g/ml of Prodrug-C respectively.^[24]

METHOD VALIDATION FOR PRODRUG-A

1A. System suitability test parameters: System suitability tests are used to verify that the resolution and repeatability of the system were adequate for the analysis intended. The parameters used in this test were the chromatographic peak resolution, theoretical plate number and tailing factor. The repeatability of these parameters was checked by solutions of Prodrug-A.^[25]

2A. Linearity and Range: Aliquots of stock solution (0.2, 0.4, 0.6, 0.8, 1.0, 1.2ml) were transferred into series of 10ml volumetric flasks and diluted up to mark with mobile phase. This yielded a solution of 20, 40, 60, 80, 100, 120µg/ml of Prodrug-A. An aliquot of 10µl of each solution was injected under operating chromatographic condition. Calibration curve of area versus respective concentration was plotted and correlation coefficient and regression line equation for Prodrug-A was calculated. Each response was an average of three determinations.^[26]

METHOD VALIDATION FOR PRODRUG-B

1B. System suitability test parameters: System suitability tests are used to verify that the resolution and repeatability of the system were adequate for the analysis intended. The parameters used in this test were the chromatographic peak resolution, theoretical plate number and tailing factor. The repeatability of these parameters was checked by solutions of Prodrug-B.^[27]

2B. Linearity and Range: Aliquots of stock solution (0.01, 0.05, 0.10, 0.15, 0.20, 0.25ml) were transferred into series of 10ml volumetric flasks and diluted up to mark with mobile phase. This yielded a solution of 1, 5, 10, 15, 20, 25µg/ml of Prodrug-B. An aliquot of 10µl of each solution was injected under operating chromatographic condition. Calibration curve of area versus respective concentration was plotted and correlation coefficient and regression line equation for Prodrug-B was calculated. Each response was an average of three determinations.^[28]

METHOD VALIDATION FOR PRODRUG-C

1C. System suitability test parameters: System suitability tests are used to verify that the resolution and repeatability of the system were adequate for the analysis intended. The parameters used in this test were the chromatographic peak resolution, theoretical plate number and tailing factor. The repeatability of these parameters was checked by solutions of Prodrug-C.^[29]

2C. Linearity and Range: Aliquots of stock solution (0.1, 0.15, 0.2, 0.25, 0.3, 0.35ml) were transferred into series of 10ml volumetric flasks and diluted up to mark with mobile phase. This yielded a solution of 10, 15, 20, 25, 30, 35µg/ml of Prodrug-C. An aliquot of 10µl of each solution was injected under operating chromatographic condition. Calibration curve of area versus respective concentration was plotted and correlation coefficient and regression line equation for Prodrug-C was calculated. Each response was an average of three determinations.^[30]

STANDARD PARACETAMOL PEAK AT 230NM IN MOBILE PHASE PHOSPHATE BUFFER: ACN (40:60V/V)



Figure-6: Chromatogram of Standard Paracetamol (R_t =2.3min) Paracetamol releases first (R_t =2.3min) because it's logP is 0.46 (highly polar).

STANDARD IBUPROFEN PEAK AT 278 NM IN MOBILE PHASE WATER: ACN (80:20V/V)



Figure-7: Chromatogram of Standard Ibuprofen (R_t =6.8min) Ibuprofen releases slow (R_t =6.8min) because it's logP is 3.97 (nonpolar).

STANDARD DICLOFENAC SODIUM PEAK AT 284NM IN MOBILE PHASE ACN:TRIETHYLAMINE BUFFER (50:50V/V)



Figure-8: Chromatogram of Standard Diclofenac Sodium (R_t=3.4min) Diclofenac sodium releases moderate (R_t=3.4min) because it's logP is 4.51 (semipolar).

Optimized Chromatographic Conditions for Prodrug-A: HPLC system: LC 2010 CHT (Shimadzu), PDA detector (PDA-SPD-M10AVP, Shimadzu), Column (Stationary Phase): Kromasil C₁₈ (150mm×4.6mm, 5µm particle size), Mobile phase : (Methanol: Water)(80:20 v/v), Flow Rate: 1.0 ml/min, Detection Wavelength: 275nm, Column oven Temp: 40°C, Run time: 10 mins. Diluent: All the final dilution of the sample was done with methanol.^[31]



Prodrug-A has logP 4.56 so release rate is higher.

Value of System suitability Parameter: Retention time (min.): 6.38, Theoretical plates: 17967.194, Tailing factor: 1.396, Resolution: 5.567, Capacity factor: 3.3.



Figure-10: Chromatogram of Prodrug-A in Acidic medium (HCl: pH=3.5) and in Alkaline medium (NaOH: pH=8.0) Paracetamol has R_t =2.5min in acidic medium and 3.5min in an alkaline medium; Ibuprofen has R_t =6.35min in acidic medium and 6.15min in alkaline medium.



Figure-11: Prodrug–A hydrolysis into Ibuprofen & Paracetamol

Method Validation:

Linearity and Range: Overlain chromatogram of Prodrug-A was shown in Figure-12. The linearity of Prodrug-A was found to be in the range of 20-120µg/ml with correlation coefficient 0.999 as shown in Figure-13.^[32]



Figure-12: Overlain Linearity Chromatogram of Prodrug-A (20-120µg/ml)



Figure-13: Calibration Curve of Prodrug-A

Precision:

1. Repeatability (Intra-day precision): The data for Intra-day precision for Prodrug-A is shown in Table-6. The %RSD for Intra-day precision was found to be 0.35-1.50 % for Prodrug-A.

Inter-day Precision (different days): The data for Inter-day Precision for Prodrug-A is shown in Table-6. The %RSD Inter-day Precision for was found to be 0.17-0.29 % for Prodrug-A.^[33]

Intra-day precision data and Inter-day precision data for estimation of Prodrug-A:

	Conc.	Mean peak	%RSD		Conc.	Mean peak	%RSD
Intra-day	(µg/ml)	Area±S.D.		Inter-day	(µg/ml)	Area±S.D.	
precision	60	1287673±6094.77	0.47	precision	60	931933±2776.952	0.29
	80	1690409±6061.18	0.35		80	1891173±3295.832	0.17
	100	2033542±68803.05	1.50		100	2694467±6749.519	0.25

*= Average of three determinations

Table-6: Comparison of Intra-day precision data and Inter-day precision data for estimation of Prodrug-A

LOD and LOQ: The Limit of Detection (LOD) was found to be 0.167μ g/ml; while the Limit of Quantification (LOQ) was found to be 0.5064μ g/ml for Prodrug-A.^[34]

Optimized Chromatographic Conditions for Prodrug-B: HPLC system: LC 2010 CHT (Shimadzu), PDA detector (PDA-SPD-M10AVP, Shimadzu), Column (Stationary Phase): Kromasil C₁₈ (150mm×4.6mm, 5µm particle size), Mobile phase : (Methanol: Water)(70:30 v/v), Flow Rate: 1.0 ml/min, Detection Wavelength: 262nm, Column oven Temp: 40°C, Run time: 15 mins, Diluent: All the final dilution of sample was done with methanol.^[35-37]



Figure-14: Chromatogram of Prodrug-B (10µg/ml) (Rt=5.5min)

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Prodrug-B has log P 4.9 so it releases slowly due to nonpolar nature.



Figure-15: Chromatogram of Prodrug-B in Acidic medium (HCl: pH=3.0) and in Alkaline medium (NaOH: pH=7.5)

Paracetamol has $R_t=2.75$ min in acidic medium and 2.8min in an alkaline medium; Diclofenac has $R_t=4$ min in acidic medium and 3.8min in alkaline medium.



Figure-16: Prodrug–B hydrolysis into Diclofenac & Paracetamol

Method Validation:

Linearity and Range: Overlain chromatogram of Prodrug-B was shown in Figure-17. The linearity of Prodrug-B was found to be in the range of 1-25 μ g/ml with correlation coefficient 0.999 as shown in Figure-18.^[35]



Figure-17: Overlain Linearity Chromatogram of Prodrug-B (1-25µg/ml)





Precision:

1. Repeatability (Intra-day precision): The data for Intra-day precision for Prodrug-B is shown in Table-7. The %RSD for Intra-day precision was found to be 0.12-0.27% for Prodrug-B.

2. Inter-day Precision (different days): The data for Inter-day Precision for Prodrug-B is shown in Table-7. The %RSD Inter-day Precision for was found to be 0.80-1.06% for Prodrug-B.^[36]

	Conc.(µg/ml)	Mean peak	%RSD		Conc.(µg/ml)	Mean peak	%RSD
Intra-day		Area±S.D.		Inter-day		Area±S.D.	
precision	10	396807±1078	0.27	precision	10	395876±3195	0.80
	15	613783±1209	0.19		15	619236±6593	1.06
	20	814533±1005	0.12		20	819518±7767	0.94

Intra-day	nrocision	hats and	Intor-day	nrecision	data for	ectimation	of Prodrug_B.
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*= Average of three determinations

Table-7: Intra-day precision data and Inter-day precision data for estimation of Prodrug-B

LOD and LOQ: The Limit of Detection (LOD) was found to be 0.023μ g/ml; while the Limit of Quantification (LOQ) was found to be 0.071μ g/ml for Prodrug-B.^[37]

Optimized Chromatographic Conditions for Prodrug-C: HPLC system: LC 2010 CHT (Shimadzu), PDA detector (PDA-SPD-M10AVP, Shimadzu), Column (Stationary Phase): Kromasil C₁₈ (150mm×4.6mm, 5µm particle size), Mobile phase : (Methanol: Water)(80:20 v/v), Flow Rate: 1.0 ml/min, Detection Wavelength: 276nm, Column oven Temp: 40°C. Run time: 10 mins. Diluent: All the final dilution of the sample was done with methanol.



Figure-19: Chromatogram of Prodrug-C (30µg/ml) (Rt=2min)



Figure-20: Chromatogram of Prodrug-C in Acidic medium (HCl: pH=3.0) and in Alkaline medium (NaOH: pH=7.5) Diclofenac has $R_t=3.4$ min in acidic medium and 3.4min in an alkaline medium; Ibuprofen has $R_t=6.8$ min in acidic medium 6.35min in alkaline medium.

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Figure-21: Prodrug–C hydrolysis into Ibuprofen & Diclofenac

Method Validation:

Linearity and Range: Overlain chromatogram of Prodrug-C was shown in Figure-22. The linearity of Prodrug-C was found to be in the range of $10-35\mu g/ml$ with correlation coefficient 0.999 as shown in Figure-23.^[38]



Figure-22: Overlain Linearity Chromatogram of Prodrug-C (11-35µg/ml)



Figure-23: Calibration curve of Prodrug-C

Precision:

1. Repeatability (Intra-day precision): The data for Intra-day precision for Prodrug-C is shown in Table-8. The %RSD for Intra-day precision was found to be 0.12-0.27% for Prodrug-C.

2. Inter-day Precision (different days): The data for Inter-day Precision for Prodrug-C is shown in Table-8. The %RSD Inter-day Precision for was found to be 0.80-1.06% for Prodrug-C.^[39]

	Conc.(µg/ml)	Mean peak	%RSD		Conc.(µg/ml)	Mean peak	%RSD
Intra-day		Area±S.D.		Inter-day		Area±S.D.	
precision	10	78678±439	0.55	precision	10	78938±742	0.94
	20	157900±1297	0.82		20	157234±1688	1.07
	30	235695±1228	0.52		30	234695±2851	1.21

*= Average of three determinations

Table-8: Intra-day precision data and Inter-day precision data for estimation of Prodrug-C

LOD and LOQ: The Limit of Detection (LOD) was found to be 0.09μ g/ml; while the Limit of Quantification (LOQ) was found to be 0.28μ g/ml for Prodrug-C.^[40]

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Value of System Suitability Parameter of Prodrug-A, Prodrug-B and Prodrug-C:

Sr.No.	Parameters	Prodrug-A	Prodrug-B	Prodrug-C
1	Retention time (min.)	6.38	5.4	1.9
2	Theoretical plates	17967.194	6696	2548
3	Tailing factor	1.396	1.04	1.2
4	Resolution	5.567	5.566	-
5	Capacity factor	3.3	2.237	-

Table-9: Value of System Suitability Parameter of Prodrug-A, Prodrug-B and Prodrug-C

Linearity data of Prodrug-A, Prodrug-B and Prodrug-C:

Pro	Conc	Peak	SD	%RSD	Pro	Conc	Peak	SD	%RS	Pro	Conc	Peak	SD	%RS
drug	(µg/	area			drug	(µg/	area		D	drug	(µg/	area		D
-A	ml)	mean*			-В	ml)	mean			-C	ml)	mean		
	20	561864	970.0	0.1726		1	49515	287	0.57		10	78656	222	0.28
	40	901074	1867.3	0.2068		5	195469	1328	0.67		15	116858	691	0.59
	60	1290471	30508.4	0.2383		10	395999	1062	0.26		20	159155	937	0.58
	80	1690463	9334.84	0.5527		15	613574	877	0.14		25	196324	1292	0.65
	100	2045532	6391.61	0.3127		20	813557	1314	0.16		30	235652	1580	0.67
	120	2476697	1051.67	0.0425		25	1003451	1297	0.12		35	274989	1483	0.53

Table-10: Linearity data of Prodrug-A, Prodrug-B and Prodrug-C

Linearity Results for Prodrug-A, Prodrug-B and Prodrug-C:

Regression Analysis	Prodrug-A	Prodrug-B	Prodrug-C	
Regression equation	Y= 19154x+15359	Y= 40261x+1949	Y= 7858x+125.5	
Correlation co-efficient	0.999	0.999	0.999	
Slope	19154	40261	7858	
Intercept	15359	1949	125.5	

Table-11: Linearity Results for Prodrug-A, Prodrug-B and Prodrug-C

Summary of Validation Parameters for HPLC method of Prodrug-A, Prodrug-B and Prodrug-C

Sr.No.	Parameters		Prodrug-A	Prodrug-B	Prodrug-C
1	Linearity Range		20–120µg/mL	1–25µg/mL	10–35µg/mL
2	Regression equation		y=19154x+15359	y=40261x+1949	y=7858x+125.5
3	Correlation co-efficient		0.999	0.999	0.999
4	Precision (%RSD)	Interday	0.17 - 0.29%	0.80-1.06%	0.94 - 1.21%
		Intraday	0.35 - 1.50%	0.12-0.27%	0.52 - 0.82%
5	Limit of Detection		0.167µg/ml	0.023µg/ml	0.09µg/ml
6	Limit of Quantification		0.5064µg/ml	0.071µg/ml	0.28µg/ml

Table-12: Summary of Validation Parameters for HPLC method of Prodrug-A, Prodrug-B and Prodrug-C





Histogram of LOD

Figure-24: LOD & LOQ [Calculated by LOD=3.3 σ /s; LOQ=10 σ /s]

CONCLUSION

Ibuprofen, Diclofenac & Paracetamol have been taken as NSAID and three Prodrugs have been synthesized by reacting of the acid chloride of ibuprofen & diclofenac with paracetamol to get Prodrug of ester linkage and an acid chloride of ibuprofen has been reacted with diclofenac to get Prodrug of amide linkage. A prodrug is a substance which after administration is metabolized into a pharmacologically active drug. Actually, Prodrug has least medicinal value in *in-vitro/in-vivo* but after biotransformation by metabolism in *in-vivo* it releases the active medicament. A drug is a substance which is a chemical entity, has a definite structural skeleton, obtained by natural or synthetic or semisynthetic source, which can fit on bioreceptor platform having controlling capacity to control over the biochemical malfunction. Every drug is xenobiotic because it is coming from the outer source (xeno) and active in biological unit (biotic). A prodrug is the precursor of drug which is made by derivatization of the same to enhance the bioavailability by pharmacokinetics, lipid solubility by partition coefficient and increase the physicochemical & biochemical parameters by pharmacodynamics. All three Prodrugs showed different log P values and molecular weights according to the solubility parameters and electronegativity: log P profile: Prodrug-C>Prodrug-B>Prodrug-A; molecular weight profile: Prodrug-C>Prodrug-B>Prodrug-A.

The main side effect of NSAID is gastric acidity due to the release of free H⁺ because all NSAIDs have free –COOH (carboxylic acid) group which act by competitive inhibition of cyclooxygenase enzyme (COX₁/COX₂). Here the target of this project has been designed in such a way to convert the free –COOH of API (ibuprofen/diclofenac/paracetamol) into Prodrug of ester (–COO–) as well as amide (–CONH–) linkage (Prodrug-A/Prodrug-B/Prodrug-C) which releases free API after metabolic hydrolysis in acidic pH: 1-4 and alkaline pH: 7-9. Since the Prodrugs are repository forms so chances to release gastric acid has been minimized due to non-availability of free –COOH group in the stomach. The biotransformation of active drug from Prodrug takes such a time in the stomach that all goes up to duodenum and then to ileum of the small intestine that chance of acidity is reduced. Finally, all Prodrugs go to the small intestine where alkaline pH starts so gastric acidity is reduced. Since all three Prodrugs are made of two NSAID: Prodrug–A (logP=4.56) releases lbuprofen & Paracetamol, Prodrug–B (logP=4.90) releases Diclofenac sodium & Paracetamol and Prodrug–C (logP=6.13) releases lbuprofen & Diclofenac which shows distinct two R_t values in HPLC both in acidic and alkaline hydrolysis and these R_t values of Prodrugs match with the individual API components so the purpose of our goal has been completed successfully. The pH of gastric acid varies from 1.5-3.5 in the human stomach lumen, the acidity being maintained by the proton pump H⁺/K⁺ ATPase. So the pattern for acid hydrolysis was adjusted at pH=3-3.5 by



HCl. The pH of intestine varies from 5.6-6.9, so the pattern for alkaline hydrolysis was adjusted at pH=7.0-8.0 by NaOH.

This project has been divided into three parts:

1. Synthesis of Prodrug of ester and amide linkages of NSAID having carboxylic acid, phenolic and imino groups

2. Logarithmic partition coefficient comparison study and molecular weight of synthesized Prodrugs of ibuprofen + paracetamol, diclofenac sodium + paracetamol and ibuprofen + diclofenac sodium 3. Study of in-vitro biotransformation of Prodrugs of

ester and amide linkages of ibuprofen, diclofenac sodium and paracetamol in acidic and alkaline medium.

Standard Rt of NSAIDs:

1. Paracetamol releases first (Rt=2.3min) because it's logP is 0.46 (highly polar). 2. Ibuprofen releases slow (Rt=6.8min) because it's logP is 3.97 (semipolar). 3. Diclofenac sodium releases moderate (Rt=3.4min) because it's logP is 4.51 (nonpolar).

Prodrug Rt after hydrolysis:

Prodrug-A [-COO-; ester linkage]: Paracetamol has Rt=2.5min in acidic medium and 3.5min in an alkaline medium; Ibuprofen has Rt=6.35min in acidic medium and 6.15min in alkaline medium.

Prodrug-B [-COO-; ester linkage]: Paracetamol has Rt=2.75min in acidic medium and 2.8min in an alkaline medium; Diclofenac has Rt=4min in acidic medium and 3.8min in alkaline medium.

Prodrug-C [-CONH-; amide linkage]: Diclofenac has Rt=3.4min in acidic medium and 3.4min in alkaline medium; Ibuprofen has Rt=6.8min in acidic medium 6.35min in alkaline medium.

Rt of paracetamol varies between 2.5-3.5min; Rt of ibuprofen varies between 6.15-6.8min and Rt of diclofenac varies between 3.4-3.8min. The focus of in-vivo metabolic profile of Prodrugs has been implemented into in-vitro hydrolytic reaction in both acidic and alkaline pH to get the satisfactory desired result.

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↓ REFERENCES

1. Jalpa G. Patel and Prof. Dr. Dhrubo Jyoti Sen; Synthesis of Prodrug of ester and amide linkages of NSAID having carboxylic acid, phenolic and imino groups: World Journal of Pharmacy and Pharmaceutical Sciences; 2016; 5(11): 897-908.

2. Dhrubo Jyoti Sen and Jalpa G. Patel; Logarithmic partition coefficient comparison study and molecular weight of synthesized Prodrugs of ibuprofen+paracetamol, diclofenac sodium+paracetamol and ibuprofen+diclofenac sodium: American Journal of Advanced Drug Delivery: 2016, 4(05), 064-068.

3. J.E. Slemmer, B.R. Martin and M.I. Damaj. Bupropion is a nicotinic antagonist. J. Pharm. Exp. Ther., 2000; 295: 321–327.

4. C. Wu, J. Quan, J. Xie, C Branford-White, L. Zhu, Y. Yu and Y. Wang. Preparation and controlled release of degradable polymeric ketoprofen-saccharide conjugates. Polym Bull., 2011; 67: 593–608.

5. M, Babazadeh and T. Mosanejhad, Vinyl ester type polymers containing ibuprofen pendents: synthesis, characterization and evaluation. Iran Polym J., 2009; 18: 179–186.

6. L. Shargel, Susanna Wu-Pong and B.C. Andrew, Applied Biopharmaceutics & Pharmacokinetics, 6th Ed.McGraw-Hill Medical Publishing Division, US., 2012; 47-66, 129-154.

7. S. Shirke^{*}, S. Shewale and M. Satpute, Prodrug design: an overview. International Journal of Pharmaceutical, Chemical and Biological Sciences., 2015; 5(1): 232-241.

8. A.V. Bhosale, G.P. Agrawal and P.Mishra: Preparation and Evaluation of Directly Compressible Forms of Mutual Prodrugs of Ibuprofen Forms of Mutual Prodrugs of Ibuprofen. Indian Journal of Pharmaceutical Sciences, 2006; 68(4): 425-431.

9. Beckett AH and Stenlake JB, Practice pharmaceutical chemistry; 4th Edn; CBS Publishers and Distributors, New Delhi, 1997, 293-304.

10. Ewing GW, Instrumental Methods of Chemical Analysis; 5th Edn; McGraw- Hill Book Company, New York, 1985, 1-7.

11. Kasture AV, Mahadik KR, Wadodkar SG and More HN, Instrumental methods of pharmaceutical analysis; 14th Edn; Nirali Prakashan, Pune, 2006, 1-30.

12. Wikipedia: The free Encyclopedia, "Ultraviolet-visible spectroscopy", 2013, http://en.wikipedia.org/wiki/Ultraviolet-visible spectroscopy.

13. Davidson AG, Practical Pharmaceutical Chemistry; 4th Edn; CBS Publishers and Distributors, 1997, 275-337. 14. Skoog DA, Holler FJ and Nieman TA, Principle of Instrumentation Analysis; 5th Edn; Thomas Asia Pvt Ltd, 2005, 580. 15. Sharma BK, Instrumental Method of Chemical Analysis; 25th Edn; Krishna Prakashan Media Ltd, 2006, 183-186. 16. Beckett AH and Stenlake JB, Practical Pharmaceutical Chemistry; 4th Edn; CBC Publication and distribution, 1977, 1-26. 17. Snyder LR., Kirkland JL and Glajch JL., Practical HPLC Method Development., 2nd Edition, Wiley Interscience, 1977, 1-26. 18. Halmilton RJ and Sewell PA., Introduction to HPLC., 2nd Edition; Chapman and Hall, 1982, 189. 19. Stability Testing of New Drugs and Product: ICH Q1 A (R2), 2003, 1-20. 20. Cartensen JT and Rhodes CT., Drug stability: Principles and practices New York – Marcel Dekker; 3rd Edition, 329 - 384. 21. Riley CM and Rosanske TW., Progress in Pharmaceutical and Biomedical Analysis., Development and validation of analytical methods., 1st Edition ; Elsevier Science Ltd, USA, 1996, 133–135. 22. Haynes, W.M. (ed.). CRC Handbook of Chemistry and Physics. 94th Edition. CRC Press LLC, Boca Raton: FL 2013-2014, p. 3-314 23. Yalkowsky, S.H., He, Yan, Jain, P. Handbook of Aqueous Solubility Data Second Edition. CRC Press, Boca Raton, FL 2010, 492. 24. O'Neil, M.J. (ed.). The Merck Index - An Encyclopedia of Chemicals, Drugs, and Biologicals. Cambridge, UK: Royal Society of Chemistry, 2013, 10. 25. American Society of Health-System Pharmacists 2013; Drug Information 2013. Bethesda, MD. 2013, 2212. 26. O'Neil, M.J. (ed.). The Merck Index - An Encyclopedia of Chemicals, Drugs, and Biologicals. 13th Edition, Whitehouse Station, NJ: Merck and Co., Inc., 2001, 876. 27. Yalkowsky SH, Dannenfelser RM; The AQUASOL database of Aqueous Solubility. Ver 5. Tucson, AZ: Univ AZ, College of Pharmacy, 1992. 28. Osol, A. (ed.). Remington's Pharmaceutical Sciences. 16th ed. Easton, Pennsylvania: Mack Publishing Co., 1980, 1057. 29. Hardman, J.G., L.E. Limbird, P.B. Molinoff, R.W. Ruddon, A.G. Goodman (eds.). Goodman and Gilman's The Pharmacological Basis of Therapeutics. 9th ed. New York, NY: McGraw-Hill, 1996, 712. 30. Testa, B. and P. Jenner. Drug Metabolism: Chemical & Biochemical Aspects. New York: Marcel Dekker, Inc., 1976, 22. 31. O'Neil, M.J. (ed.). The Merck Index - An Encyclopedia of Chemicals, Drugs, and Biologicals. Whitehouse Station, NJ: Merck and Co., Inc., 2006, 522. 32. Finn A et al; Acta Technol Legis Med, 1986, 4: 33-44. 33. American Society of Health-System Pharmacists 2011; Drug Information 2011. Bethesda, MD. 2011, 2113. 34. Anojo Hetal; Eur J Drug Metab Pharmacokinetic, 1999, 24(4): 345-51. 35. Hosale AV, Agrawal GP and Mishra P: Preparation and characterization of mutual Prodrugs of ibuprofen. IJPSR 2004; Vol. 2, Issue 4 ISSN: 0975-8232. 36. Koren, K., Mann, A.R., Colosimo, A.L., Diaz, Z., Nuclelman, A., Levovich, I., Jing, Y., Waxman, S. and Miller, W.H. Jr., Mol. Cancer Res., 2003, 1, 903. 37. Slemmer J.E., Martin B.R., Damaj M.I. Bupropion is a nicotinic antagonist. J. Pharm. Exp. Ther. 2000; 295: 321-327. 38. Wu C, Quan J, Xie J, Branford-White C, Zhu L, Yu Y, Wang Y. Preparation and controlled release of degradable polymeric ketoprofen-saccharide conjugates. Polym Bull. 2011; 67: 593–608. 39. Gennaro, R., Alfanso, W., Remington: The science and practice of pharmacy, Lippincott Williams & Wilkins, 2003, 20, 1, 913-914. 40. Shargel AB and Andrew BC, Applied Biopharmaceutics & Pharmacokinetics, 4th Ed. McGraw-Hill Medical Publishing Division, US. 47-66, 129-154. Vol. 5, Issue 8 | magazine.pharmatutor.org